

# A MICROFLUIDIC APPROACH TO THE FORMULATION OF CATIONIC AND NEUTRAL LIPID NANOPARTICLES LOADING AND PROTECTING pDNA: TOWARDS LINKING BENCHTOP METHODS AND INDUSTRIAL SCALABILITY FOR THE PRODUCTION OF GENE THERAPEUTICS

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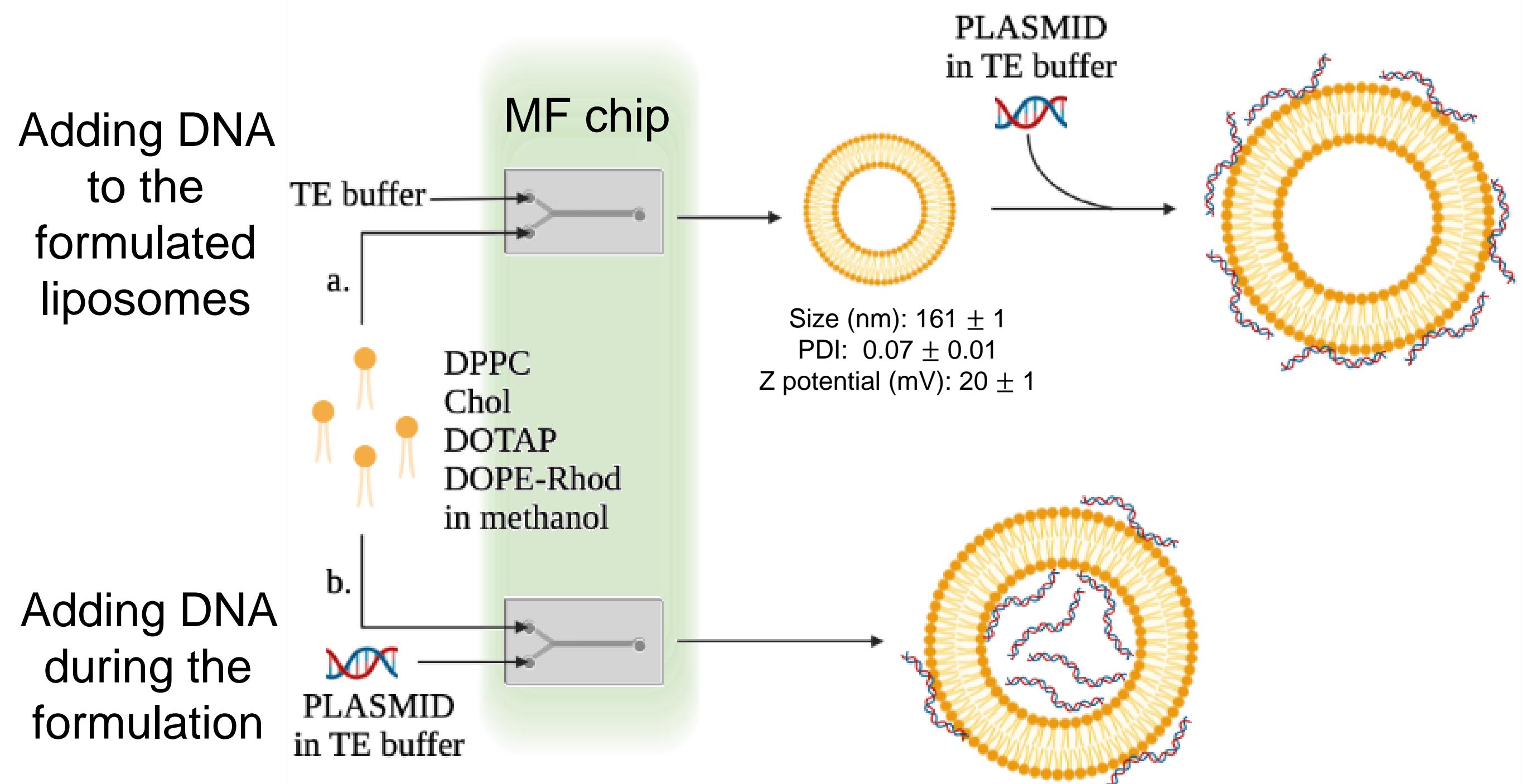
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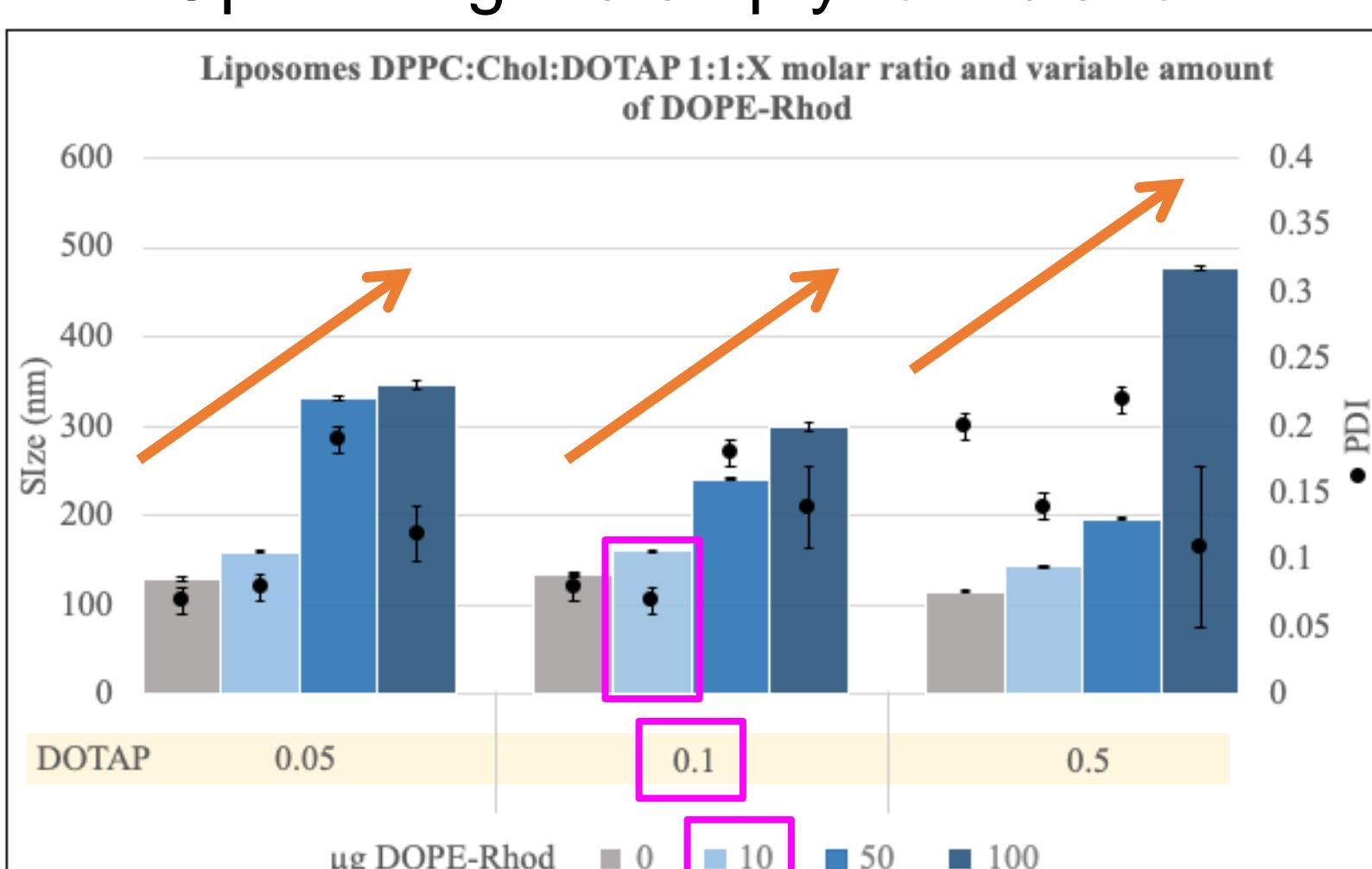
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We formulated cationic liposomes with a microfluidic chip to load eGFP plasmid DNA using two methods:



## Optimizing the empty formulation

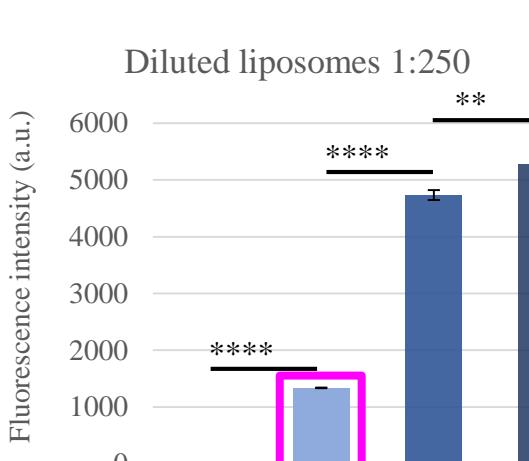


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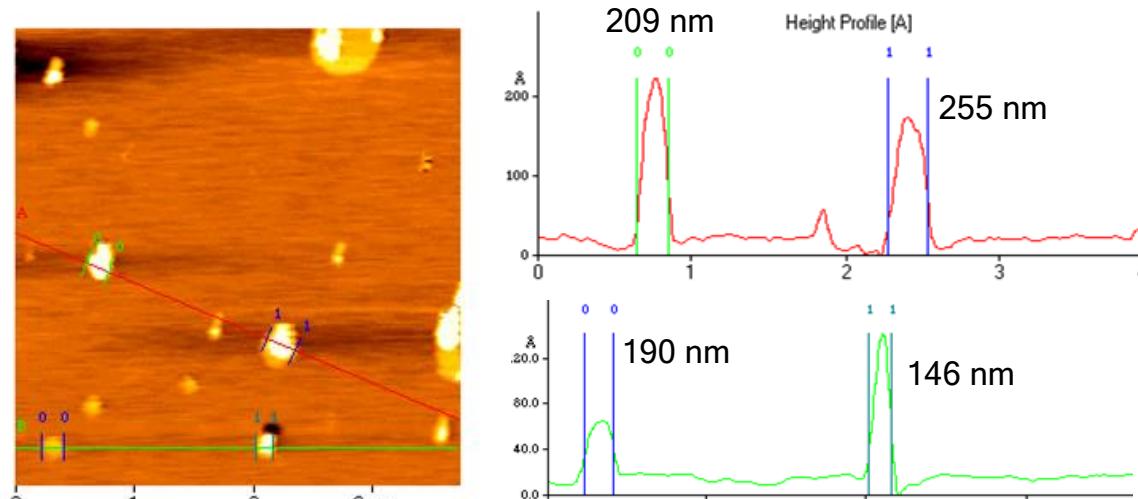


Stable in size and PDI when stored at 4°C for up to 14 days

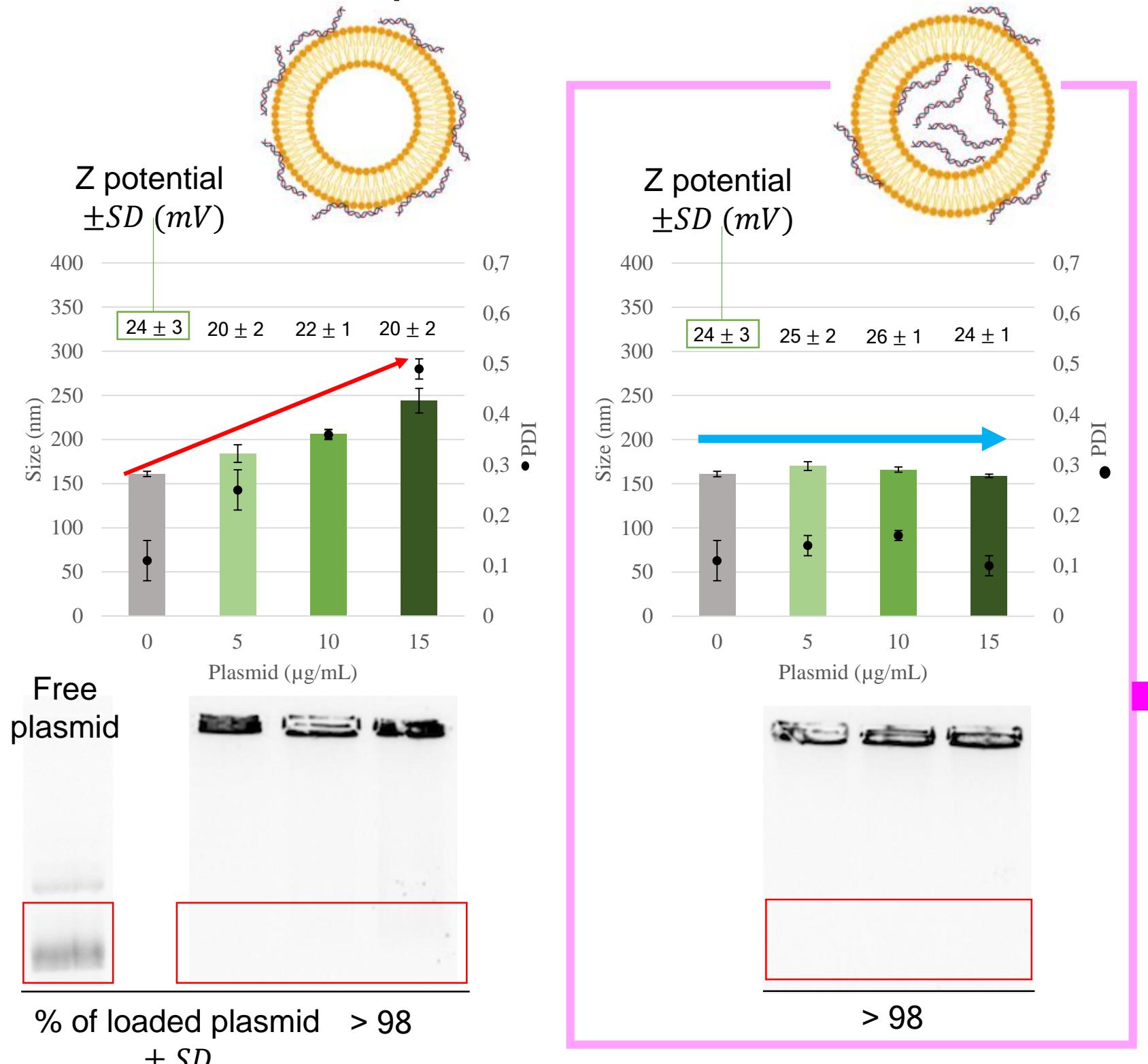
## Fluorescence intensity of Liposomes with 0-10-50-100 µg DOPE-Rhod



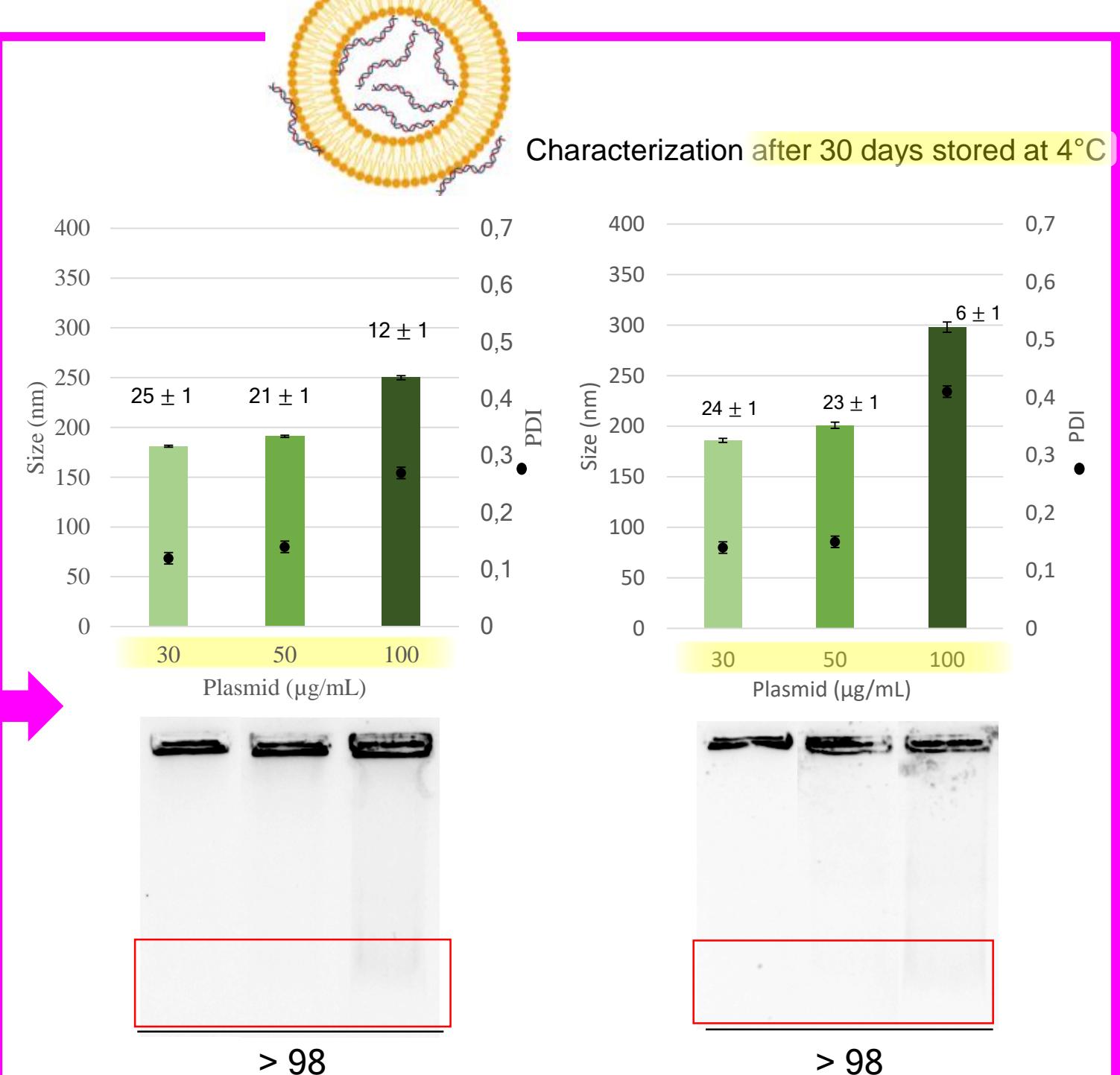
## AFM morphology of empty liposomes



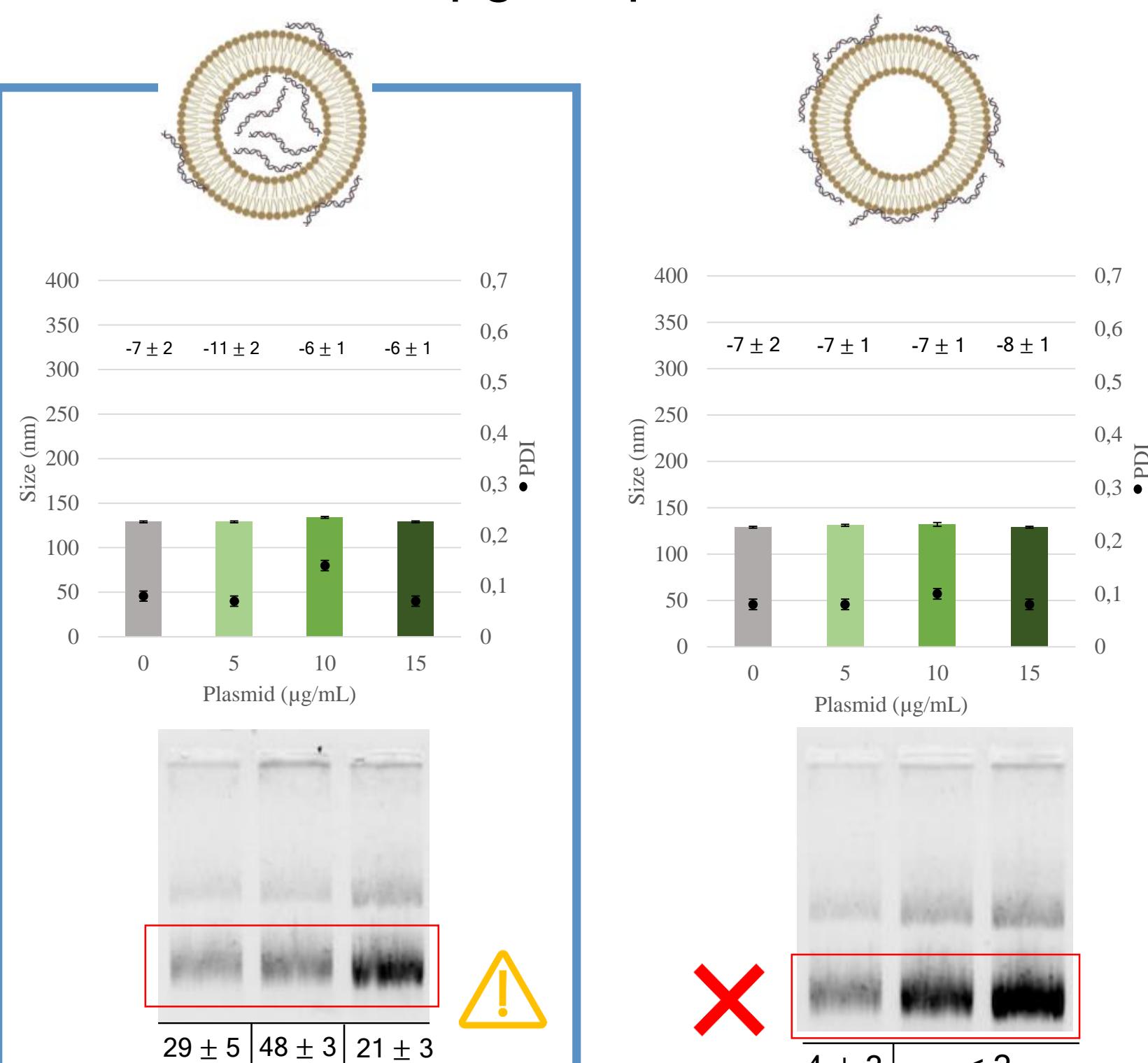
## Optimized cationic liposomes loaded with 5, 10, or 15 µg/mL pDNA



## Optimized cationic liposomes loaded with 30, 50, or 100 µg/mL pDNA + storage stability at 4°C



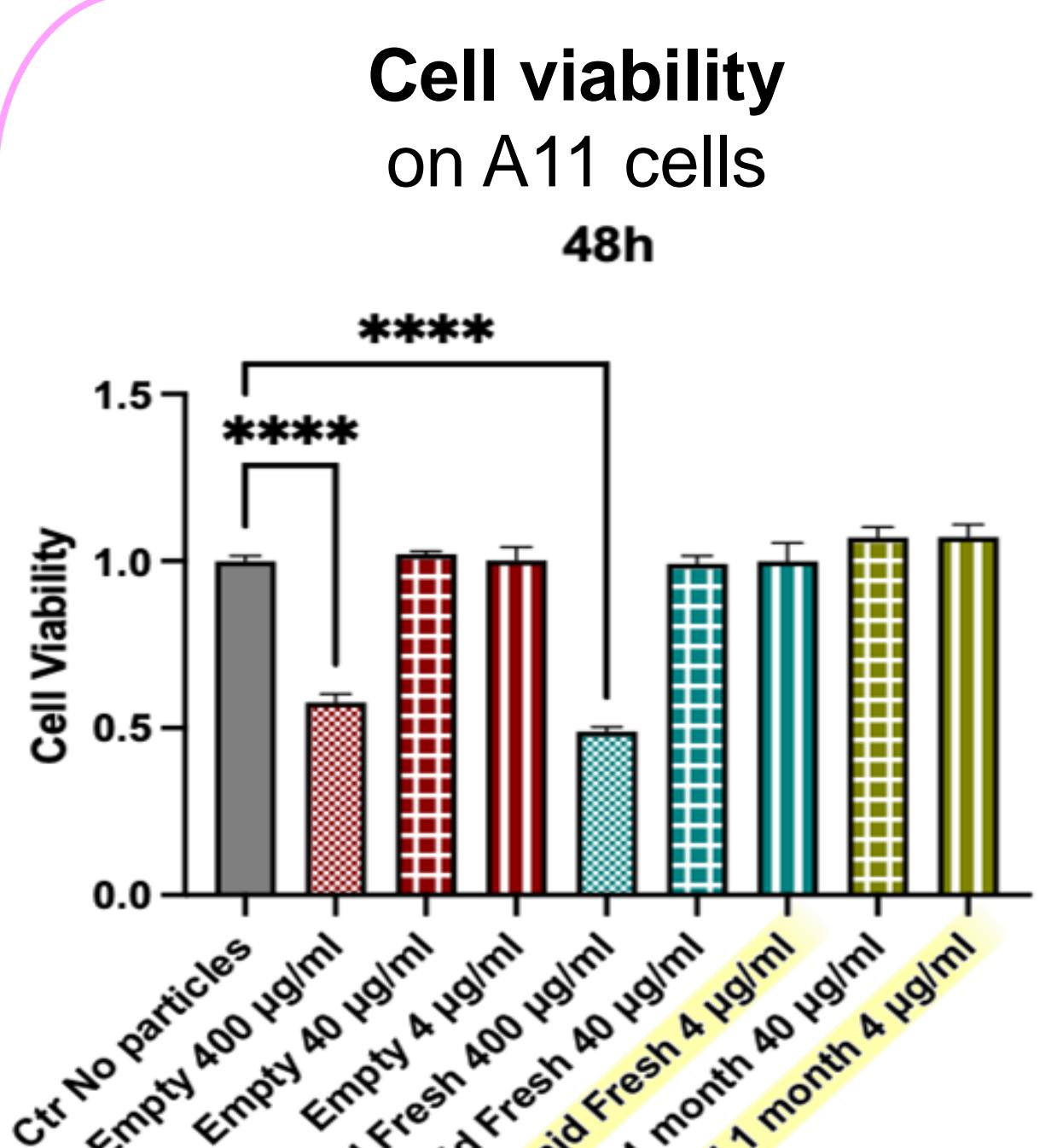
## Neutral liposomes prepared with both methods, adding 5, 10, 15 µg/mL pDNA



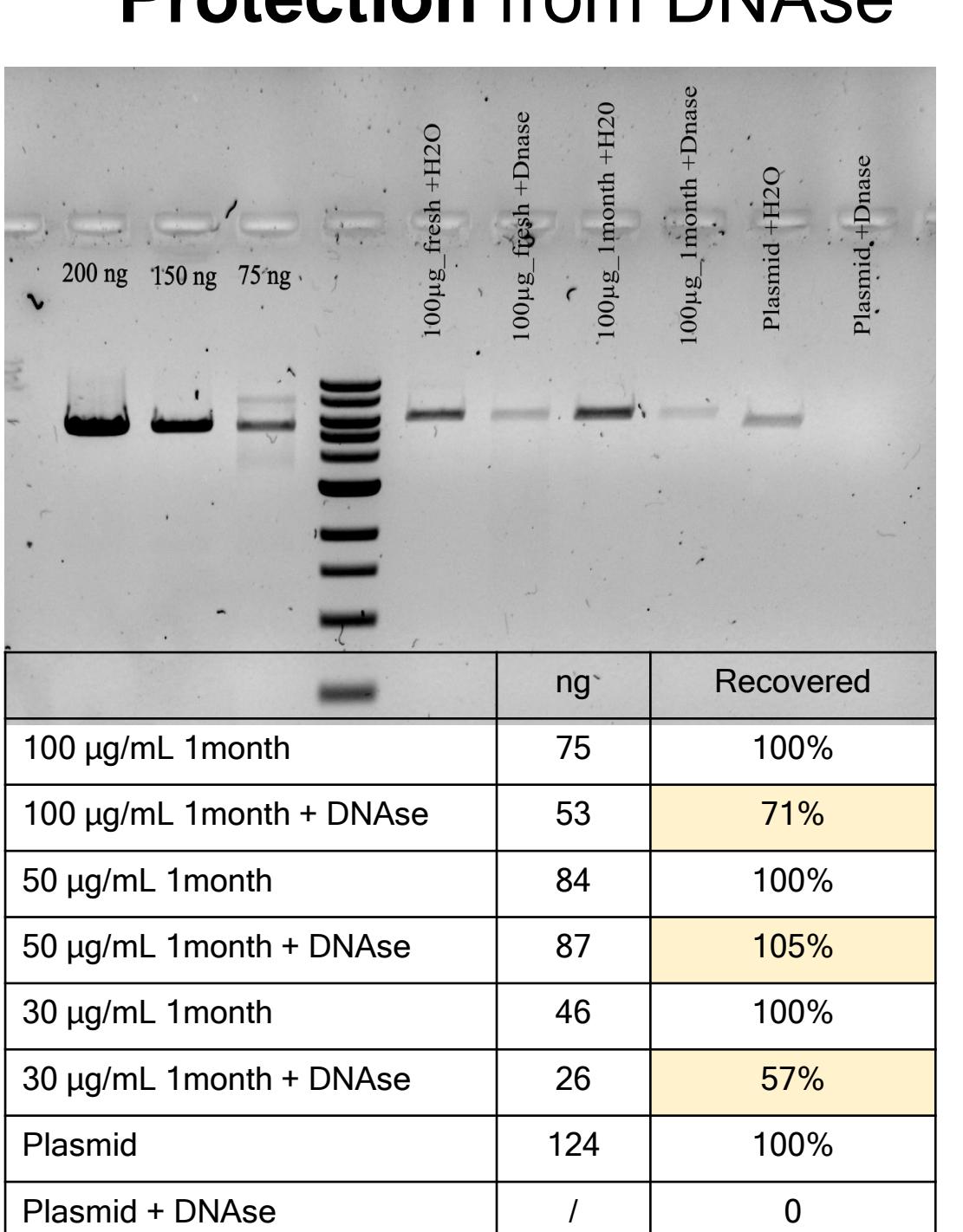
- Both methods allowed for a full **complexation of > 98% pDNA**
- Addition of pDNA to formed liposomes caused an **increase in size** and PDI dependant on the amount of DNA added

- Almost **100% of pDNA** at the highest concentration of **100 µg/mL** was loaded into cationic liposomes
- Size, PDI, and content are **stable for 1 month** at 4°C

- Adding pDNA to formulated **neutral liposomes** resulted in < 2% loading due to unspecific bindings
- Up to **50% of pDNA** was loaded when added **during the formulation of neutral liposomes**

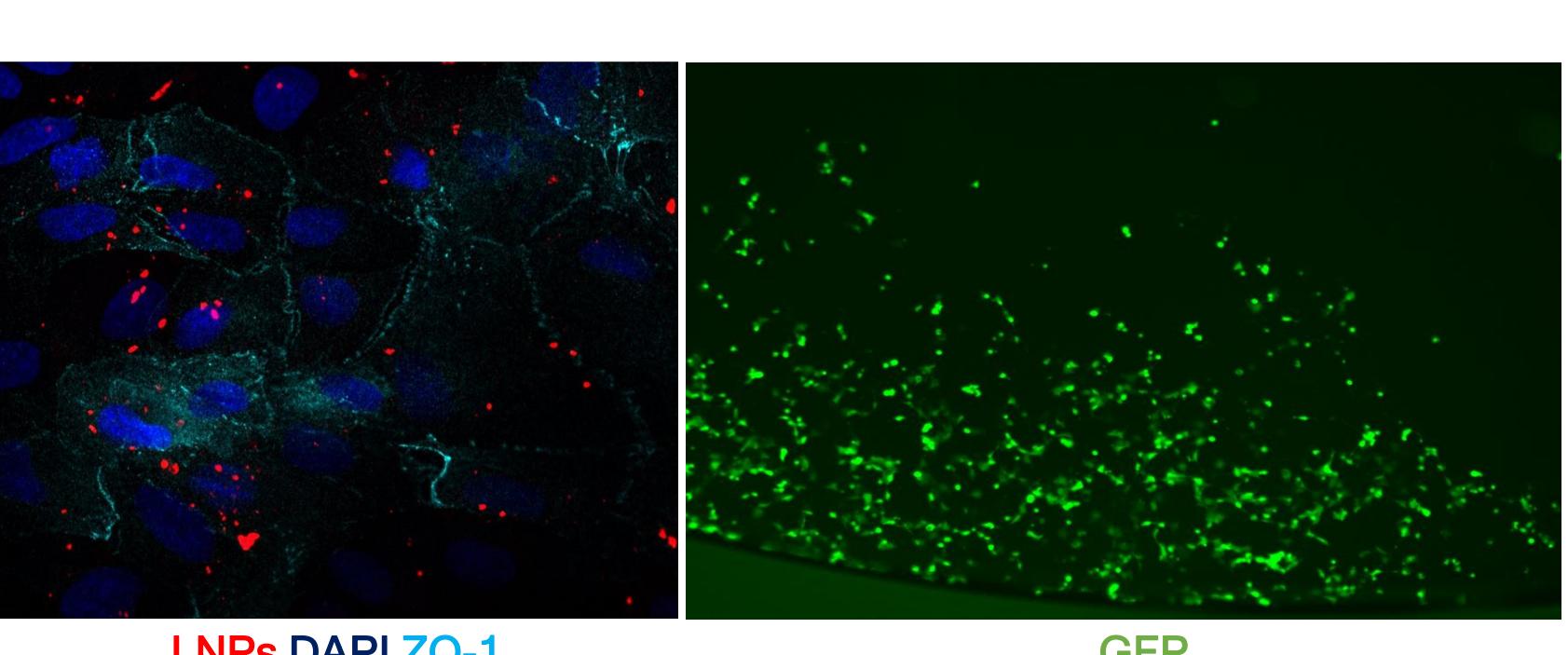


## Protection from DNase



## Transfection ability *in vitro* of LNPs loaded with 100 µg/mL pDNA

- In COS7 cells there was **no transfection** from DNA-LNPs (left)
- DNA was extracted and used with metafectene (right) showing that the DNA is still functional
- The DNA is **not released from the LNPs**



- Concentrations < 40 µg/mL of fresh or 1 month stored Liposomes showed no toxicity
- Exposure to DNase showed 30% loss of DNA

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